

Ecology and Transmission Dynamics of Visceral Leishmaniasis in Ethiopia

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Characterization of sand fly breeding sites in vertisols in North-Western Ethiopia

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Introduction

Phlebotomine sand flies are the principal vectors of Leishmania Ross, 1903 (Kinetoplastida: Trypanosomatidae), a group of protozoan parasites infecting populations exceeding 12 million worldwide During recent years foci of VL have been described in the Sheraro district near villages surrounded by vertisol fields. The successful identification and characterization of breeding and resting sites of leishmaniasis vectors in vertisol in North East Ethiopia may provide essential information for the development of measures for their control

Results

Vast majority of sand flies trapped belonged to the genus Sergentomyia (2568/2592). Significantly more Sergentomyia



Fig. 1 – Cracked vertisol – Putative breeding site of phlebotomine sand flies

spp. were captured in emergence traps placed under trees than in open fields (514±180 and 64 ± 29 sand flies/trap/night respectively), P=0.002 (Fig 4). Initial readings taken inside he soil at the end of rainy season showed stable temperature and humidity. Oxygen concentrations in shaded areas (0.7±13.5%) differed significantly from those in open fields (19.19±0.6%, P>0.0001) (Fig. 5). Subterranean temperature readings in shaded areas were stable and lower (26°C) than open fields (28°C). The relative humidity in subterranean habitats stabilized at 100% in all habitats. Microclimatic conditions were measured repeatedly during early and mid dry season. While Oxygen concentration had stabilized around atmospheric values, temperatures in deep cracks decreased with increased depth. Mean volumetric water content was high in both habitats reaching 17.79±2.77% under tree canopies, and 20.37±1.21% in open fields. Volumetric water content increased significantly with the increase in depth (P< 0.0001, fig. 6). Organic matter content reached 5.38±0.14% under tree canopy, and 5.49±0.08% in open field. However, the difference was insignificant for different habitats (P= 0.579) and different depths (P=0.950) (Fig. 7).



Fig. 6 - Volumetric water content at two habitats

and four sampling depths

Objectives

- \succ To identify putative sand fly breeding sites in vertisols in rural areas in NW Ethiopia.
- > To characterize the ambient conditions inside vertisol cracks including microclimate and physico/chemical parameters.

Materials and Methods

Putative breeding sites in cracked vertislol were covered with sandfly proof net . Emerging sand flies were trapped on large sticky traps placed inside the net. Traps were baited with incandescence light or fermenting sugar solution (Fig 2). Temperature, relative humidity and soil volumetric water content were monitored continuously in two habitats: area shaded by trees canopy and open vertisol field.

Air samples were drawn and oxygen concentrations were measured at two different depths during low and high sand fly season. . Soil samples were analyzed for organic mater, pH, and electrical conductivity. DNA was extracted from blood-fed sand fly females and the blood host were determined by PCR and sequencing of the *cyt b* gene.

Fig. 4 - Sand fly trapped at putative vertisol habitats



Fig. 5 - Oxygen Concentration at two habitatand two sempling depth



Blood engorged females were tested for host blood meal (n=90). Human blood was identified in 9 Phlebotomus orientalis (putative vector of VL in the region). Human and bovine or camel blood were identified in 6 *Phlebotomus* orientalis . Interestingly, human blood was also identified in 34 of the 43 engorged Sergentomyia spp. females (79%).



Fig. 8– PCR amplification with cytB primer (left side) and RLB (right side) of Sergentomyia spp. sand flies





Discussion

The trapping of phlebotomine sand flies emerging from cracked vertisols supports our assumptions that vertisols serve as sand fly breeding sites. Furthermore, the microclimatic conditions inside cracks were stable with high relative humidity and soil moisture throughout the year, making them suitable for development of sand flies. Larval food may be provided by decomposing organic matter, demonstrated by both rich organic matter content and the low oxygen concentrations sampled during rainy seasons. The low oxygen concentration may be caused by microbial metabolic activity depleting oxygen. The proximity to villages apparently provides human and live-stock blood sources for questing females.